ISSN: 2231-5101

## Regular Article Effects of heavy metals on *Eleusine coracana* (L.) Gaertn

### Suman Krishania\* and Kalpana Agarwal

Department of Life Sciences, International College for girls, The IIS university, SFS
Mansarovar, Jaipur(Raj) India
\*Corresponding author e-mail: Biotech.suman@gmail.com

Heavy metal (Abiotic Factor) pollution is one of the most troublesome environmental problems faced by mankind nowadays. Lead and Nickel in particular, pose serious problems due to its widespread industrial and agricultural use. Plants in general are very sensitive to Pb and Ni toxicity, displaying metabolic disturbances and growth inhibition. Eleusine coracana (L) Gaertn (var. PR202) which mainly grows in dry condition faces the toxicity of Pb and Ni which may have negative effect on its yield. Some inorganic nutrients can overcome the toxicity of heavy metals and addition of these inorganic nutrients in media can increase the resistance (tolerance level) of plant against heavy metal toxicity. Embryogenic sectors of seed callus from control (MS + 2 mg/l 2,4-D + 0.5 mg/l Kn) were sub cultured on regeneration medium (MS + 1mg/l NAA) supplemented with of toxic level of heavy metals and varied concentrations of inorganic nutrients (MgSO<sub>4</sub> and ZnSO<sub>4</sub>) to minimize the toxicity of heavy metals. Callus induction obtained only in the medium containing 100 µM concentrations of Pb and Ni but there was better growth obtained in Ni containing medium than Pb. These results show that Pb and Ni above 100 µM concentration were toxic for callus induction in Eleusine coracana but Pb was more toxic than Ni and same results was observed during plant regeneration. It was concluded that ZnSO<sub>4</sub> at the four times of MS level best to minimize the toxicity of Nickel and MgSO4 at three times of MS level best to minimize the toxicity of Lead.

**Keywords**-Heavy metals, Lead, Nickel, Magnesium Sulphate, Zinc Sulphate.

Eleusine coracana (var. PR202 45) was used as a model cereal crop. Looking to the scope and realizing its importance, the present study undertaken was investigate the effects of different concentrations of Pb and Ni elements on callus induction and plant regeneration following Objectivesstandardize a medium with growth regulators for tissue culture of Eleusine coracana. To study the influence of Pb and Ni in the morphogenesis of *Eleusine coracana* tissue culture. To optimize the culture condition with an objective of minimizing Pb and Ni toxicity and maximizing the yield .To Implantation in field to increase the productivity of cereals. Levels of

different heavy metals were varied taking MS as the standard medium. Today Abiotic stress is a major global problem limiting crop productivity. Stress factors are a serious problem limiting the yield potential modern cultivars. Stress nutritional imbalances in the plant causing reduction in water uptake and toxicity, decreasing the production. Abiotic stresses like salinity, heavy metals and pesticides are the primary cause of crop failures in India. It has been estimated that about 8.6 million ha of land is affected by salinity in India (Pathak, 2000). Stresses are increasing drastically today because of pollution, declining availability of quality water and land degradation. Ragi is used as cereal,

which is also called finger millet and scientific name is *Eleusine coracana*. *Eleusine* coracana belonging to family Poaceae. Cereals constitute a major source of food for the human population of the world. Important cereals in day to day use are rice, wheat and millets. Of all the millets Eleusine contains more percentage of different nutrients as compared to rice and wheat. It is a rich source of calcium, magnesium and potassium. Eleusine is nutritionally very rich cereal and is a staple food for poor and invalids (Panda, 1999). Due its nutritional superiority and requirement by poor people production needs to be improved. Eleusine coracana mainly growing in dry condition faces heat, salinity and heavy metal stress which may cause negative effect on its yield. Stress tolerant plants can also be developed by breeding and by transgenesis which are complex processes. (Babu et al, 2007) Tissue culture techniques offer an easy and important tool in developing stress tolerant variants (Nabors and Dykes, 1985.). Finger millet contains more fiber, minerals and vitamins, which are normally deficient in the Indian diet, and has eight times more calcium than other cereals The high calcium, high soluble fiber, low fat and low glycemic index of malted grains is effective in controlling the blood glucose levels of diabetics. Studies were conducted on diabetics (male and female) living in different rural and urban locations in 2008 in different countries (Adikant prahan et al, 2008). Finger millet is especially valuable as it contains amino acid methionine, which is lacking in the diets of millions of the poor people. Finger millet can be ground and cooked into cakes, puddings or porridge. The grain is made into a fermented drink (or beer) in many parts of Africa. The straw from finger millet is used as animal fodder. It is also used for as a flavored drink in It has been shown that the festivals. nutrient level in the medium has profound effect on callus induction and subsequent plant regeneration (Ramage and Williams 2002). There also occurs interplay between nutrients and plant growth regulators

(PGRs) during the course of in vitro growth differentiation (DeFossard 1974). Appropriate levels of nutrients may also partially substitute the requirement of PGRs in the plant regeneration medium (Preece 1995; Poddar et al. 1997). Bregitzer et al. (1998, 2000) and Dahleen and Bregitzer (2002) investigated the effect of various micronutrients on barley culture and reported several fold better regeneration of green plants by combining the various improvements. Optimized levels of copper lead to improved plant regeneration from callus cultures of barley (Bregitzer et al. 1998; Castillo et al. 1998; Chauhan and Kothari 2004), rice (Sahrawat and Chand 1999), finger millet (Kothariet al. 2004), sorghum (Nirwan and Kothari 2003) and wheat (Tahiliani and Kothari 2004). Sahasrabudhe et al. (1999), found an increase in the embryogenic response of indica rice on higher concentrations of boric acid. These nutrient levels which were adequate for tobacco tissue culture may not necessarily be optimum for the culture of other plant species like graminaceous monocots (Dahleen 1995).

### **Materials and Methods**

An important plant *Eleusine corcana* (L.) Gaertn of family Poaceae was taken as the model plant for the study (Fig 2) Seeds of agronomically superior and released variety (PR 202) of ragi were procured from Agricultural University, Bangalore (Fig1) and this study was conducted in 2011 at IIS University Department of Life Sciences, Jaipur.





Figure 1. (Left) Seeds of *Eleusine coracana*, Figure 2. (Right) Mature Plant of *Eleusine coracana* in field.

Common Name: Fingermillet Vernacular Name: Ragi Explants Taken: Seeds

### Sterilization and Preparation of Explants

Explants taken were aseptically sterilized. Sterilization process of seeds was carried out in the laminar airflow cabinet. Seeds were surface sterilized in 0.1 percent mercuric chloride solution for 3 minutes. These explants (seeds) were inoculated on MS medium supplemented with auxin 2, 4-D (2 mg/l) and cytokinin Kn (0.5 mg/l).

### **Basal Medium Preparation**

Basal medium used in the present study was MS (Murashige and Skoog, 1962) medium. Sucrose: 3% (w/v), Agar: 0.8-1% (w/v) and pH: 5.8.

### Aseptic Manipulations

Aseptic culture was carried out in laminar air flow chamber.

### Incubation

During the entire work, cultures were incubated in growth chamber equipped with two air conditioners and temperature controller to maintain temperature at26±1°c. A photoperiod of 16 hours alternating with 8 hours of darkness was maintained.

# Preparation of Medium with Varied Concentration of Heavy Metals (Lead and Nickel)

All the constituents of the MS medium i.e. inorganic and organic nutrients, growth regulators and sucrose were mixed in required amount and the Volume of medium was raised by adding distilled water to attain the final concentration of nutrients in the medium as per requirement. The medium was poured in beakers and then the different amounts of heavy metals (the concentration of which is to be varied i.e. Pb or Ni) were added separately in each beaker. The beakers were marked for the concentration of heavy metal (Table 1-2).

### **Statistical Analysis**

The observations recorded for the various experiments were subjected to following statistical analysis. The average (mean) was calculated by dividing the sum of values of observations for a particular treatment by the total number of observations for that treatment.

### **Standard Deviation**

This is a measure of dispersion which was calculated by squaring the deviation of each observation from the mean, adding the squares, dividing by the number of observation and extracting the square.

Table 1- Callus induction medium for E. coracana with various concentrations of lead.

Medium	Concentration of Pb (µM)
MS + 2,4-D + Kn (Control)	Nil
MS + 2,4-D + Kn+Pb	$100~\mu\mathrm{M}$
MS + 2,4-D + Kn +Pb	300 μΜ
MS + 2,4-D + Kn+Pb	500 μM

Growth regulators added: Auxin- 2, 4D, Cytokine- Kinetin, Agar - 8gms/liter.

Table 2- Callus induction medium for *E. coracana* with various concentration of nickel.

Medium	Concentration of Ni (μM)
MS + 2,4-D + Kn(Control)	Nil
MS +2,4-D + Kn + (NiSO4)	$100~\mu\mathrm{M}$
$MS + 2,4-D + Kn+(NiSO_4)$	300 μΜ
MS + 2,4-D + Kn+( NiSO <sub>4</sub> )	500 μΜ

### **Steps of Experiment -1)**

**Primary Callus Induction A)**Seeds was inoculated on MS Medium supplemented with 2mg/l 2,4-D+0.5mg/l Kinetin (control) After 3-4weeks induced callus of the control seeds were inoculated on MS medium with lower concentration of 2,4-D(0.2mg/l).

- B) Seeds was inoculated on MS Medium supplemented with 2mg/l 2, 4-D+0.5mg/l Kinetin (control) In addition to this in one set of experiment lead concentration were varied in callus induction medium and in second set Nickel was varied, the amount of callus formed in each case was record and after 3-4 weeks all the calli were transferred to (MS+1mg/l NAA) regeneration media. Approximately 4-5 seeds were cultured in each flask and approx 5 replica of each concentration were made.
- **2)** Embryogenic callus cultures and Plant Regeneration: Approx 250mg of callus was transferred to each flask for regeneration. The embryogenic callus obtained from primary subculture control, as well as the variation of Pb/Ni were transferred on regeneration i.e.MS+1mg/l NAA media.
- 3) Effect of Inorganic Nutrient in Overcoming Toxicity Third set of experiment was planned to study effect of

some inorganic nutrients in overcoming the toxicity of Pb and Ni. Out of all these concentration of Pb/Ni toxic level was chosen.

Now regeneration media was prepared which was MS+NAA and addition of only nontolerant concentration of Pb/Ni with various concentration of inorganic nutrients (MgSO<sub>4</sub>,ZnSO<sub>4</sub>).

On this media embryogenic callus obtained from control after 3-4 subculture, approx 250 mg of callus(control) was transferred to each flask of regeneration media and addition of toxic concentration of Pb/Ni (300µM) with various concentration (0x, 1/2x, 1x, 2x, 3x, 4x) of inorganic nutrients (MgSO<sub>4</sub>ZnSO<sub>4</sub>).

#### **Results**

A) Seed Culture (Table 3)After 2-3 weeks of inoculation of seeds, it was observed that the callus so developed was healthy, compact, nodulated, darkgreen embryogenic sectors along with watery translucent non embryogenic sectors (Fig 3a). Amount of callus formed was recorded (Fig 3b).Only the embryogenic sectors were transferred to maintenance medium; the non embryogenic portion of the callus was excluded from being subculture.

Table 3. Callus formation from seeds of *Eleusine coracana* L. cultured on MS medium Supplemented with 2, 4-D and Kn.

Media Amount of callus formed per seed(mg)	
<u> </u>	
250	
296	

C.I.M=Callus Induction Medium, C=Control,S.D= Standard deviation R.M=Regeneration Medium and M.M=Maintenance Medium

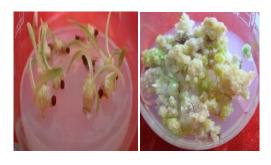


Figure 3. Response of seeds on Callus Induction Medium and Response after this Primary callus was subculture on 2, 4-D (0.2 mg/l). a. Callus induced from seeds of *Eleusine coracana* on MS + 2, 4-D (2 mg/l) +Kn (0.5 mg/l). b. Embryogenic callus after subculture on MS medium with 2, 4-D (0.2 mg/l).

**B)** Callus Culture- Experiments for evaluating the effects of different heavy metals (Pb and Ni) were conducted in three sets. In the first set, seeds were used as the plant material and the heavy metal concentration was varied in callus induction medium (MS + 2mg/1 2, 4-D and 0.5 mg/1 Kn). These calli were then transferred to regeneration medium (MS + 1mg/1 NAA). In the second set of experiments primary callus was subcultured on maintenance medium (MS + 0. 2 mg/1 2, 4-D) and then

subcultured on regeneration medium (MS+1mg/lNAA) supplemented with varied concentrations of heavy metals. In the third set of experimental embryogenic sectors of primary callus from control (MS + 2 mg/l 2,4-D + 0.5 mg/l Kn)were sub cultured on regeneration medium (MS + 1mg/l NAA) supplemented with toxic concentration of heavy metals and varied concentrations of nutrients (MgSO<sub>4</sub>,ZnSO<sub>4</sub>) (Fig3a-b).

Table 4. Variation in callus induction medium with varied concentration of lead regeneration response

Concentrations of Pb in Callus Induction Medium (µM)	Amount of callus formed / explants (mg)	Number of shoots per seed callus on plant regeneration medium ± S.D.
0 μΜ <sup>c</sup>	250	5.4 ±2.1
100μΜ	198	3.3±0.4
300μΜ	0	0
500μΜ	0	0

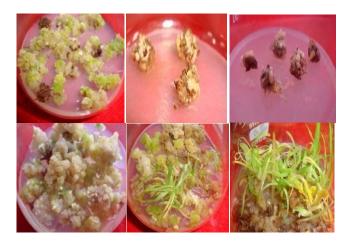


Figure 4. Effects of different concentrations of Pb on primary callus induction and plant Regeneration in *Eleusine coracana* as compare to control. a) MS+ (2 mg/l) 2, 4-D and (0.5 mg/l) kinetin +Lead (100 $\mu$ M); b) MS+(2 mg/l) 2, 4-D and (0.5 mg/l) kinetin +Lead (300 $\mu$ M); c)MS+ (2 mg/l) 2, 4-D and (0.5 mg/l) kinetin +Lead (500 $\mu$ M); d)MS+ (2 mg/l) 2, 4-D and (0.5 mg/l) Kinetin (control) e) Primary callus obtained on callus induction Medium with (100 $\mu$ M) Pb when sub cultured on MS medium with 1mg/l NAA (Regeneration medium); f) Primary callus sub cultured on MS medium with1mg/NAA(control).

Effects of Heavy Metals on Callus Induction and Plant Regeneration (lead and nickel)

### 1. Effects of Pb on primary callus induction and plant regeneration

In *Eleusine coracana*, regeneration was recorded via formation of shoot buds on

enlarged apicaldomes and meristematic nodules (Kumar et al.2001). Here seeds of *Eleusine coracana* were inoculated on modified callus induction medium (2 mg/l 2, 4-D and 0.5 mg/l kinetin) with varied Pb levels (100 $\mu$ M, 300  $\mu$ M and 500  $\mu$ M). Calli were formed in treatments with 100  $\mu$ M

concentration of lead-nodulated, compact, embryogenic and soft, watery non embryogenic callus. The primary callus from  $100~\mu\text{M}$  concentration of lead was then transferred to normal regeneration medium (MS + 1mg/l NAA). After 2-3 weeks deep green and well developed regenerates appeared (Fig 4a-f) (Table 4).

### 2. Effects of NiSO<sub>4</sub> on primary callus induction and plant regeneration

Seeds of *Eleusine coracana* were inoculated on modified callus induction medium (2 mg/l 2, 4-D and 0.5 mg/l kinetin) with varied NiSO<sub>4</sub> and levels (100 $\mu$ M, 300  $\mu$ M and 500  $\mu$ M). After about 2-3 weeks of incubation, two types of calli were formed

in treatments with 100 µM concentration of nickel- nodulated, compact, embryogenic and soft, watery non embryogenic callus. While there was no callus formation in treatments with 300 and 500 μM concentrations of nickel. Callus growth was better on nickel containing medium than lead containing medium. The primary callus from 100 µM concentration of nickel was then transferred to normal regeneration medium (MS + 1mg/1 NAA). After 2-3 weeks of sub culture, deep green and well developed regenerants appeared. Nickel was considered less toxic than lead but callus and regeneration was worse than control. (Fig 5a-f) (Table 5).

Table 5. Variation in callus induction with varied concentrations of Nickel and regeneration response

Concentrations of NiSO <sub>4</sub> in Amount of callus formed Number of shoots per seed callus on			
Callus Induction Medium(µM) / explants (mg)		Plant Regeneration Medium ± S.D	
0 μΜ <sup>c</sup>	250	5.2 ± 1.4	
100μΜ	206	$3.8 \pm 0.7$	
300μΜ	0	0	
500μΜ	0	0	



Figure 5. Effects of different concentrations of NiSO<sub>4</sub> on primary callus induction and plant regeneration in *Eleusine coracana* as compare to control. a) MS+(2 mg/l) 2, 4-D and (0.5 mg/l) kinetin +Nickel 100 $\mu$ M; b) MS+(2 mg/l) 2, 4-D and (0.5 mg/l) kinetin +Nickel300 $\mu$ M; c)MS+(2 mg/l) 2, 4-D and (0.5 mg/l) kinetin +Nickel500 $\mu$ M; d) MS+(2 mg/l) 2,4-D and (0.5 mg/l) kinetin. e) Primary callus obtained on callus induction medium with (100 $\mu$ M) NiSO<sub>4</sub> when subcultured on MS medium with 1mg/l NAA (Regeneration medium). f) Primary callus sub cultured on MS medium with 1mg/l NAA (control)

### 3. Effects of Pb on Plant Regeneration

Embryogenic callus from maintenance medium (MS + 0. 2 mg/l 2,4-D) were further transferred on regeneration medium (MS + 1mg/l NAA) supplemented with varied concentrations of Pb (100μM, 300 μM and 500 μM) to see the effects of Pb concentrations on

regeneration of *Eleusine coracana*. After 2-3 weeks, differentiation of shoot buds took place in medium containing 100µM

concentration of Pb. Number of shoots per callus piece was counted (Fig 6a-d) (Table6).

Table 6. Embryogenic callus (MS+2, 4-D 0.2mg/l) response in variation in plant regeneration medium with different concentration of Lead.

Concentrations of Pb	Mean number of shoot formed/Callus	
in plant Regeneration Medium (μM)	Piece ± S.D.	
0 μΜ <sup>c</sup>	5.2 ± 1.1	
$100 \mu M$	$3.8 \pm 2.2$	
300μΜ	0	
500μΜ	0	

Initial weight of callus=296mg



Figure 6. Effects of Pb variations on Plant Regeneration from embryogenic callus.

a) Plant regeneration from embryogenic callus on MS + NAA containing with Pb (100μM).

b) Plant regeneration from embryogenic callus on MS + NAA containing with Pb (300μM).

c)Plant regeneration from embryogenic callus on MS + NAA containing with Pb (500μM)

d)Plant regeneration from embryogenic callus on MS + NAA containing without Pb

Table7. Embryogenic callus (MS+2, 4-D 0.2mg/l) response in variation in plant regeneration medium with different concentration of Nickel sulphate.

Concentrations of NiSO <sub>4</sub> in plant Regeneration Medium(μM)	Mean number of shoots formed/ callus piece ± S.D
Control	5.6 ± 1.7
100μΜ	$4.2 \pm 0.9$
300μΜ	0
500μΜ	0



Figure 7) Effects of NiSO<sub>4</sub> variations on Plant Regeneration from embryogenic callus. a)Plant regeneration from embryogenic callus on MS+ NAA containing with NiSO<sub>4</sub> (100 $\mu$ M). b)Plant regeneration from embryogenic callus on MS+NAA containing with NiSO<sub>4</sub> (300 $\mu$ M). c)Plant regeneration from embryogenic callus on MS+ NAA containing with NiSO<sub>4</sub> (500 $\mu$ M). d)Plant regeneration from embryogenic callus on MS+NAA containing without NiSO<sub>4</sub> (control).

4. Effects of NiSO<sub>4</sub> on Plant Regeneration Embryogenic callus from maintenance medium (MS + 0. 2 mg/l 2,4-D) were further transferred on regeneration medium (MS + 1mg/l NAA) supplemented with varied concentrations of NiSO<sub>4</sub> (100 $\mu$ M, 300  $\mu$ M and 500  $\mu$ M)After 2-3 weeks, differentiation of shoot buds took place in medium containing 100 $\mu$ M concentration of NiSO<sub>4</sub> and number of shoots per callus piece was counted (Fig7a-d) (Table7).

### Effects of inorganic nutrients on Lead and Nickel toxicity

### 5. Effects of MgSO<sub>4</sub> on lead toxicity for Plant Regeneration

It was observed that MgSO<sub>4</sub> at 2x concentration of the MS medium was able

for overcoming lead toxicity. The callus induced on callus induction medium with 100µM Pb when cultured on regeneration medium with MS level of magnesium produced 3.3 shoots per callus piece. When 300 µM Pb was increased to two times the normal level of MS medium MgSO<sub>4</sub> 4.2 shoots were obtained per callus piece suggesting that MgSO<sub>4</sub> at this level can overcome lead toxicity. This number was almost similar to regeneration under control conditions i.e. callus induced on normal callus induction medium cultured on normal regeneration medium. However it was seen that on increase of the MgSO4 to four times of normal level improved regeneration (Fig8a-f) (Table 8).

Table 8. Variation in plant regeneration, response from embryogenic callus of *Eleusine coracana* on MS with toxic level of Pb (300µM) and varied concentrations of MgSO<sub>4</sub>

· · ·	,	
The toxic level of Pb in	Concentrations of MgSO <sub>4</sub> (mM)	Mean number of shoots
Regeneration Medium	added in Regeneration Medium	formed/callus ±S.D
	0mM	0
	0.75mM	0
<b>300</b> μM	1.5* mM	0
	3mM	$4.2 \pm 0.9$
	4.5mM	5.2 ± 1.1
	6mM	$5.6 \pm 0.4$

<sup>\*</sup> Normal level in the ms medium and initial weight of callus=250mg



Figure 8. Effects of MgSO<sub>4</sub> on Lead toxicity in Plant Regeneration. a)Plant regeneration on MS medium without MgSO<sub>4</sub>+NAA (1mg/l). b) Plant regeneration on MS medium with 0.75mM MgSO<sub>4</sub>+NAA (1mg/l). c) Plant regeneration on MS medium with 1.5 mMMgSO<sub>4</sub> +NAA (1mg/l). d) Plant regeneration on MS medium with 3mM MgSO<sub>4</sub>+NAA (1mg/l). e) Plant regeneration on MS medium with 4.5mMMgSO<sub>4</sub> +NAA (1mg/l). f) Plant regeneration on MS medium with 6mM MgSO<sub>4</sub>+NAA (1mg/l).

### 6. Effects of ZnSO<sub>4</sub> on Nickel toxicity for Plant Regeneration

It was observed that ZnSO<sub>4</sub> at 2x concentration of the MS medium was able for overcoming nickel toxicity. The callus induced on callus induction medium with  $100\mu M$  NiSO<sub>4</sub> and when subcultured on regeneration medium with MS level of magnesium produced 4.2 shoots per callus piece. When ZnSO<sub>4</sub> was increased to two

times the normal level of MS medium 4.9 shoots were obtained in regeneration medium. However it was seen that on increase of the ZnSO<sub>4</sub> to three times of normal level improved regeneration but further increase of ZnSO<sub>4</sub> at Four times decrease the regeneration frequency (Fig 9a-f) (Table 9).

Table 9. Variation in plant regeneration response from embryogenic callus of *Eleusine corcana* on MS with toxic level of NiSO<sub>4</sub> (300 $\mu$ M) and varied concentrations of ZnSO<sub>4</sub>.

	Concentrations of ZnSO <sub>4</sub> (mM)	Mean number of shoots
Regeneration Medium	added in Regeneration Medium	formed/explant ±S.D
	0mM	0
	14.95mM	0
300μΜ	29.91*mM	0
	59.82mM	$4.9 \pm 1.7$
	89.73mM	$5.4 \pm 0.8$
	119.64µM	$4.4 \pm 0.5$



Figure 9) Effects of ZnSO<sub>4</sub> on Nickel toxicity in Plant Regeneration. a) Plant regeneration on MS medium without ZnSO<sub>4</sub> +NAA (1mg/l). b)Plant regeneration on MS medium14.95mM ZnSO<sub>4</sub>+NAA (1mg/l). c) Plant regeneration on MS medium 29.91\*mM ZnSO<sub>4</sub>+NAA (1mg/l). d) Plant regeneration on MS medium 59.82mM ZnSO<sub>4</sub>+NAA (1mg/l). e) Plant regeneration on MS medium 89.73mM ZnSO<sub>4</sub>+NAA (1mg/l). f) Plant regeneration on MS medium 119.64 $\mu$ M ZnSO<sub>4</sub>+NAA (1mg/l).

### Discussion

Biotechnology of millets has lagged behind the major cereal crops due to difficulties in plant regeneration and poor transformation efficiencies (Kothari et al. 2005). Inorganic nutrients are major components of MS medium and hence offer to be the best variable to study their effect on the morphogenetic potential of the plant under study. Of all the micronutrients, Magnesium and Zinc has gained utmost importance because it is known to enhance the embryogenic callus growth, and plant regeneration efficiency in several other plants (Popelka and Altpeter 2001; Nirwan and Kothari 2003; Tahiliani and Kothari 2004; Joshi and Kothari 2007). According to Ramage and Williams (2002) in some cases callus proliferation and plant regeneration can be improved by modifying the salt concentration, irrespective of the growth regulator used, the regenerants grown from such a medium performed better in Eleusine and many times there is increase in regeneration response in the presence of elevated levels of Mg and Zn was observed. This observation was supported by various workers in different plants (Dahleen 1995; Bregitzer et al. 1998; Yang et al. 1999; Zhang et al. 1999; Chauhan and Kothari 2004; Joshi and Kothari 2007). In our study an improvement of in vitro response has been discerned when inorganic nutrients was added to the MS medium. However, Witte et al. (2002) completely denied the role of CoCl2 in plant tissue culture. They reported presence of CoCl2 to accumulation of urea which is not good for plant growth. Similarly, higher levels of CoCl2 also stimulated the production of betalains (Trejo-Tapia et al. 2003), which inhibits morphogenesis in both rice and barley at concentrations higher than the optimum. The increasing world population and the unpredictable changes in the world climate pose a threat on the already shrunken food basket. Millets are an excellent alternative cereal for coeliac patients. Coeliac disease is a lifelong intolerance to glutin found in wheat, barley and rye, the prevalence rate of which is found to be 1:100 (Mc Gough and Cummings 2005). Improvement in the millet yield and production is attempting alternate. Various studies with manipulated levels of mineral proved that every species has its particular level of mineral requirement. Catering the need of a particular plant further requires the extensive evaluation of tissue culture medium to optimize the regeneration response.In Eleusine coracana (L.) Gaertn which mainly grows in dry condition faces

the toxicity of Lead and Nickle, which may have negative effect on its yield and these heavy metals decrease the growth rate of explant in in vitro conditions. Some inorganic nutrients can overcome the toxicity of heavy metals. At non-toxic concentration of heavy metals, addition of these inorganic nutrients in media can increase the resistance of explant against heavy metal toxicity. Experiments for evaluating the effects of different heavy metals (Pb and Ni) were conducted in three sets. Callus induction was obtained only in the medium containing 100 µM concentrations of Pb and Ni but there was better growth obtained in Ni containing medium than Pb. These results show that Pb and Ni above 100 µM concentration were toxic for callus induction in Eleusine coracana but Pb was more toxic than Ni. Embryogenic sectors of the seed callus obtained from media containing 100 µM concentration of Pb and Ni was sub cultured in the regeneration medium (MS + 1 mg/ 1 NAA) for shoot regeneration. Sub cultured regenerants appear deep green and well developed. Sub culturing of callus obtained from control maintenance medium (MS + 0. 2 mg/l 2,4-D ) on regeneration medium (MS + 1mg/l NAA) supplemented with varied concentrations of Pb and Ni also resulted in shoot regeneration in medium containing 100 µM concentration of Pb and Ni. These results show that Pb and Ni were also toxic for shoot regeneration in *Eleusine* coracana above 100 µM concentration. Embryogenic sectors of seed callus from control (MS + 2 mg/l 2,4-D + 0.5 mg/l Kn) were sub cultured on regeneration medium (MS + 1mg/l NAA) supplemented with of toxic level of heavy metals and varied concentrations of inorganic nutrients (ZnSO<sub>4</sub> and MgSO<sub>4</sub>) to minimize the toxicity of heavy metals. Results obtained show that ZnSO<sub>4</sub> at the four times of MS level was best to minimize the toxicity of lead and MgSO4 at the three times of MS level was best to minimize the toxicity of nickel.

#### Conclusion

experiments These led to conclusion that lead and nickel above 100 uM concentration are inhibitory for callus well for plantlet induction as as regeneration in Eleusine coacana. It can be concluded that magnesium and zinc can minimize the toxicity of Pb and Ni but Zinc at four times of MS level was resulted worse shoot induction three times.

### Acknowledgment

My heartfelt thanks to Dr. Kalpana Agarwal Associate professor, Botany Department, International College for girls, Jaipur, THE IIS UNIVERSITY Jaipur-20 in 2011.

#### References

- Adikant Pradhan., S. K. Nag and Pati 2010. Dietary management of finger millet (*Eleusine coracana* L. Gaerth) controls diabetes Current Science, 98, 6-25.
- Babu S, Sheeba A, Yogameenakshi J and P Rangaswamy (2007). Effect of salt stress in the selection of salt tolerant hybrids in rice (*Oryza sativa* L.) under *in vivo* and *in vitro* conditions. Asn J Plt Sci 6:137-142.
- Bregitzer P, Campbell RD, Dahleen LS, LemauxPG,ChoM-J(2000) Development of transformation systems for elite barley cultivars. Barley Genet News 30:1-4
- Bregitzer P, Dahleen LS, Campbell RD (1998) Enhancement of plant regeneration from embryogenic callus of commercial barley cultivars. Plant Cell Rep 17:941–945
- Castillo AM, Egana B, Sanz JM, Cistue L (1998)Somatic embryogenesis and plant regeneration from barley cultivars grown in Spain. Plant Cell Rep 17:902–906
- Chauhan M, Kothari SL (2004) Optimization of nutrient levels in the medium increases the efficiency of callus induction and plant regeneration in recalcitrant Indian barley (*Hordeum*

- *vulgare* L.) in vitro. In Vitro Cell Dev Biol Plant 40:520–527
- DahleenLS (1995) Improved plant regeneration from barley callus cultures by increased copper levels. Plant Cell Tissue Organ Cult 43:267–269
- Dahleen LS, Bregitzer P (2002) An improved media system for higher regeneration rates from barley immature embryo derived callus cultures of commercial cultivars. Crop Sci 42:934–938
- DeFossard RA (1974) Responses of the callus from zygotal and microsporal tobacco (*Nicotiana tabacum* L.) to various combinations of iodole acetic acid and kinetine. New Phytol 77:699
- Joshi A, Kothari SL (2007) High cooper levels in the medium improves shoot bud differentiation and elongation from the cultured cotyledons of *Capsicum annuum* L. Plant Cell Tissue Organ Cult 88:127–133
- Kothari SL, Agrawal K, Kumar S (2004) Inorganic nutrient manipulation for highly improved in vitro plant regeneration in finger millet [*Eleusine coracana* (L.) Gaertn]. In Vitro Dev Biol Plant 40:515–519
- Kothari SL, Kumar S, Vishnoi RK, Kothari A, Watanabe KN (2005) Applications of biotechnology for improvement of millet crops: review of progress and future prospects. Plant Biotechnol 22:81–88
- Kumar S, Agarwal K, Kothari SL (2001) In vitro induction and enlargement of apical domes and formation of multiple shoots in finger millet, *Eleusine coracana* (L.) Gaertn and crowfoot grass, *Eleusine indica* (L.) Gaertn. Curr Sci8:1482–148
- Mc Gough N, Cummings JH (2005) Coeliac disease: a diverse clinical syndrome caused by intolerance of wheat, barley and rye. Proc Nutr Soc 64:434–450
- Nabors M W and T A Dykes (1985). Tissue culture of cereal cultivars with increased salt, drought and acid tolerance. Biotech in international agri res, IRRI Phillipines, pp: 121-138.

- Nirwan RS, Kothari SL (2003) High copper levels improve callus induction and plant regeneration in Sorghum bicolor (L.) Moench. In Vitro Cell Dev Biol Plant 39:161–164
- Panda P R, (1999). Incorporating millets in high intensity cropping. Yojana.
- Pathak (2000). A tool for arresting land degradation- Indian Farming. Agro forestry 49:15-19 Plant Tissue Culture Biotech: 26-36.
- Poddar K, Vishnoi RK, Kothari S (1997)
  Plant regeneration from embryogenic callus of finger millet *Eleusine coracana* (L.) Gaertn. on higher concentrations of NH4NO4 as a replacement of NAA in the medium. Plant Sci 129:101–106.
- Popelka JE, Altpeter F (2001) Interactions between genotypes and culture media components for improved in vitro response of rye (Secale cereale L.) inbred lines. Plant Cell Rep 20:575–582
- Preece JE (1995) Can nutrient salts partially substitute for plant growth regulators. Plant Tissue Cult Biotechnol 1:26–37
- Ramage CM, Williams RR (2002) Mineral nutrition and plant morphogenesis. In Vitro Cell Dev Biol Plant 38:116–124
- Sahasrabudhe NA, Nandi M, Bahulikar RA (1999) Influence of boric acid on somatic embryogenesis of a cytosterile line of indica rice. Plant Cell Tissue Organ Cult 58:73–75
- Sahrawat AK, Chand S (1999) Stimulatory effect of copper on plant regeneration

- in indica rice (*Oryza sativa* L.). J Plant Physiol 154:517–522
- Tahiliani S, Kothari SL (2004) Increased copper content of the medium improves plant regeneration from immature embryo derived callus of wheat (*Triticum aestivum*). J Plant Biochem Biotechnol 13:85–88
- Trejo-Tapia G, Arias-Castro C, Rodriguez MM (2003) Influence of the culture medium constituents and inoculums size on the accumulation of blue pigment and cell growth of Lavandula spica. Plant Cell Tissue Organ Cult 72:7–12
- Witte PC, Tiller SA, Taylor MA, Davies HV (2002) Addition of nickel to Murashige and Skoog in plant culture activates urease and may reduce metabolic stress. Plant Cell Tissue Organ Cult 68:103-104
- Yang YS, Zheng YD, Chen YL, Jain YY (1999)Improvement of plant regeneration from long term cultured calluses of Tipei-309, a model rice variety in in vitro studies. Plant Cell Tissue Organ Cult 57:199–206
- Zhang S, Cho MJ, Koprek T, Yun R, Bregitzer P, Lemaux PG (1999) Genetic transformation of commercial cultivars of oat and barley using in vitro shoot meristematic cultures derived from germinating seedlings. Plant Cell Rep 18:959–966.