



ISSN: 2075-6240

# Phytochemical profile and antioxidant potential of leaf and bark extracts of *Cassine glauca* (Rottb.) Kuntze

K. Dhanasekaran¹, Kishore Kumar Ashok², K. Palanisamy¹, P. Subramaniam¹, V. Balakrishnan¹\*

<sup>1</sup>PG and Research Department of Botany, Arignar Anna Government Arts College, Namakkal-637002, Tamil Nadu, India, <sup>2</sup>Integrative Agriculture Department, College of Agriculture and Veterinary Medicine, United Arab Emirates University, P.O.Box No.15551, Al Ain, United Arab Emirates

#### **ABSTRACT**

In the present study the phytochemical composition and antioxidant prospective of Cassine glauca leaf and bark extracts. Leaf and bark of C. glauca were extracted by methanol and chloroform through Soxhlet extraction. The extracts were analyzed for qualitative and quantitative phytochemical constituents, including phenolics, flavonoids, tannins, and other bioactive compounds. Antioxidant activity was determined through five different methods such as DPPH radical scavenging, reducing power, assay of molybdenum, hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) scavenging, and nitric oxide scavenging assays. Phytochemical analysis revealed that methanol extracts from both leaf and bark were more effective than chloroform, showing higher concentrations of phenolics, tannins and flavonoids. The leaf extract had a total phenol content of 47.458 mg GAE/g, tannins at 45.298 mg RE/g. and a flavonoid content of 160.106 mg TAE/g, and the bark extract showed lower but notable levels of these compounds. The leaf extract showed greater antioxidant activity when compared with bark extract in all experiments, includes DPPH radical scavenging activity, molybdenum assay, reducing power activity, nitric oxide scavenging activity and H<sub>2</sub>O<sub>2</sub> scavenging activities. Methanol extracts of C. glauca leaf and bark exhibited significant antioxidant properties, with the leaf extract showed greater efficacy. The present study highlights a prospective source of C. glauca as a potential resource of natural antioxidants and suggests additional exploration of its therapeutic benefits in oxidative stress-related disorders. The results emphasize the effectiveness of methanol as a solvent for extracting potential bioactive compounds from C. glauca.

Received: November 10, 2024 Revised: April 29, 2025 Accepted: May 01, 2025 Published: May 12, 2025

\*Corresponding author:
V. Balakrishnan
E-mail: palanivbalu@gmail.com

KEYWORDS: Cassine glauca, Antioxidants, Phytochemicals, Therapeutics, Bioactive compounds, Efficacy

#### **INTRODUCTION**

Traditional medicine has become increasingly important worldwide in recent years, particularly in developing and developed nations where numerous people depend on medicinal plants for their basic health care needs (Mosihuzzaman, 2012). Historical records emphasize the crucial role of herbal medicine specialists in promoting well-being (Iyiola & Wahab, 2023). The healing properties of medicinal herbs are linked to their various phytochemical compounds, including flavonoids, phenols, alkaloids, and terpenes (Raina et al., 2014). Factors like population growth, limited synthetic pharmaceuticals, high costs, and adverse effects of man-made medications drive increased use of plant-based medicines. Phytomedicine involves using phytochemical extracts or phytocompounds to interact with cells, tissues, or organs to manage conditions such as diabetes, and cardiovascular diseases, and prevent oxidants, and infections (Hassan, 2012). Plant secondary metabolites are good potential antimicrobial agent and derived from different parts of the herbal plants (Sundari & Kavitha, 2024).

Medicinal plants are defined as plants with therapeutic value in any part such as leaves, stems, bark, roots, flowers, or fruits are employed in the creation of medicinal mixtures or as ingredients in industrialized pharmaceutical drugs. These plants are frequently utilized in traditional or alternative and complementary medicine for the prevention and treatment of particular illnesses and conditions (Chintamunnee & Mahomoodally, 2012). They may be located as wild species naturally present in terrestrial ecosystems with minimal human disturbance or cultivated domestically through selective breeding for commercial purposes, providing bioactive compounds essential for pharmaceutical drug production (Calixto, 2000).

Addressing oxidative stress with medicinal plants involves enhancing bioavailability, understanding interactions,

Copyright: © The authors. This article is open access and licensed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.o/) which permits unrestricted, use, distribution and reproduction in any medium, or format for any purpose, even commercially provided the work is properly cited. Attribution — You must give appropriate credit, provide a link to the license, and indicate if changes were made.

personalizing treatments, ensuring safety, and integrating with conventional medicine (Muscolo et al., 2024). By adopting a comprehensive approach, it is possible to harness the benefits of medicinal plants while overcoming challenges related to drug resistance and efficacy. Phytochemical compounds were identified through various sophisticated instrumentation methods such as HPLC, GC-MS and LC-MS (Kavitha et al., 2024). Various phytomolecules, including quercetin are help to prevent the oxidative damage in human beings by modulating glutathione levels, enzyme activity, and signal transduction pathways (Xu et al., 2019). To date, numerous bioactive compounds have been discovered and characterized from the medicinal plants that have been effectively utilized for biomedical applications. Several phytocompounds derived from natural products are currently undergoing clinical trials. Despite the increasing interest in conventional medicine, the demand for alternative medicine remains strong, prompting ongoing research in this field. Numerous chemical compounds produced by medicinal plants exist as valuable secondary metabolites (Dhanasekaran et al., 2025).

Cassine is a notable tropical genus that belongs to the Caesalpiniaceae family comprising around 600 species around the globe. C. glauca is an indigenous to India, tropical Asia and Australia. This species is distinguished by its smooth, evergreen characteristics and its swift growth into shrubs or small trees that can reach heights of 2 to 5 meters. The leaves are bipinnately compound, showcasing 4 to 6 pairs of ovate leaflets with notched tips. Its flowers are vibrant yellow and are found in axillary racemes. Historically, C. glauca has been utilized for numerous medicinal applications (Albrahim et al., 2021). Previous research has examined the antioxidant potential of various extracts derived from the aerial parts and seeds of C. glauca (Veerapur et al., 2017). Nonetheless, there is a scarcity of studies focused on the antioxidant properties of the leaves and bark of C. glauca. Hence, the current study aimed to perform qualitative and quantitative phytochemical analysis and to identify the active compounds present in this plant and explore the antioxidative characteristics of C. glauca to tackle oxidative stress, aiming to better understand its potential therapeutic benefits against oxidative damage.

#### **MATERIALS AND METHODS**

#### Collection of Plant materials

Leaves and bark of *Cassine glauca* (Rottb.) Kuntze were collected in Namakkal, identified with the help of plant taxonomist and its specimen sample was deposited at the Herbarium of PG and Research and Department of Botany, Arignar Anna Government Arts College, Namakkal (#NKL2019/12). The collected plant samples were shade dried, powdered and used for extraction.

# **Preparation of Solvent Extracts**

Fine powdered plant samples (Leaf and bark) (20 g) were packed into the thimble of the extraction apparatus, which was placed in the Soxhlet tube and subjected to an 8-hour extraction process using heat. Two separate extractions were performed,

one with 250 mL of methanol and another with chloroform. The concentrated vapors obtained from the extracts were further processed and dehydrated using a Rotary Vacuum Evaporator (Equitron, India) at a temperature of 60 °C under reduced pressure. The solvent was carefully evaporated, and the resulting dried crude residues were aseptically weighed, the extract value was then calculated using the formula: Weight of the dried extract/Weight of the plant material×100. The extracts were then re-dissolved in their respective solvents and stored at 4 °C in sterile, labeled, airtight containers for subsequent analysis. This meticulous process ensured the preservation and concentration of the plant extracts for subsequent investigations (Sreelatha & Padma, 2009).

# **Phytochemical Analysis**

The leaf and bark extracts of *C. glauca* underwent qualitative analysis to detect flavonoids, alkaloids, phenolics, glycosides, tannins, coumarins, sugars, proteins, terpenoids, and amino acids. The methodology followed the procedures outlined by Harborne (1973) and Kokate (2005).

#### **Phenolic Content**

To assess the total phenolic content of the extracts, Folin-Ciocalteu reagent method was employed, and gallic acid was used as a reference standard. Solutions of gallic acid standards (ranging from 0.01 to 0.05 mg/mL) as well as plant extracts (0.1 and 1 mg/mL) were prepared with methanol. In each tube, 0.5 mL of either the standard or the extract was combined with 2.5 mL of diluted Folin-Ciocalteu reagent and 2 mL of a 7.5% sodium carbonate solution. After allowing the mixture to sit for at least 30 minutes at room temperature, the absorbance was measured at 760 nm using a spectrophotometer. The intensity of the blue color, which correlates to the phenolic content, was reported as mg of gallic acid equivalents (GAE) per gram of the extract. Measurements were conducted in triplicate to ensure accuracy.

## **Tannin Content**

To assess the tannin levels in the leaf and bark extracts of *C. glauca*, the technique utilizing insoluble polyvinylpolypyrrolidone (PVPP) was used as described. One milliliter of each extract, prepared in methanol at a concentration of 1 mg/mL, was combined with 100 mg of PVPP. The mixture was subjected to vortexing, followed by incubation at 4 °C for 15 minutes, and then centrifuged at 3,000 rpm for 10 minutes. The resulting clear solution, which contains non-tannin phenolics, was evaluated for total phenolic content using a similar method as was applied for the total phenolics. The tannin content was determined as the difference between the total phenolic content and the non-tannin phenolic content.

#### Flavonoid Content

The total flavonoid content in the leaf and bark extracts of C. glauca, followed the method outlined. For each extract, 1 mL of the sample was combined with 4 mL of distilled water and

 $0.30~\mathrm{mL}$  of a  $10\%~\mathrm{NaNO}_2$  solution. After 5 minutes,  $0.30~\mathrm{mL}$  of a  $10\%~\mathrm{AlCl}_3$  solution was introduced, followed by the addition of 2.0 mL of a  $1\%~\mathrm{NaOH}$  solution. The mixture was mixed thoroughly, and the absorbance was recorded at 510 nm using a blank for comparison. The results were reported as quercetin equivalents (mg quercetin/g dried extract) derived from a standard curve for quercetin (0-12 mg/mL).

# Antioxidant activity of Leaf and Bark Extracts of C. glauca

Based on the phytochemical study, methanol extracts from the leaves and bark of C. glauca were selected for the antioxidant activity viz. DPPH radical scavenging assay, molybdenum content assay, Reducing power assay, nitric oxide assay and hydrogen peroxide  $(H_2O_2)$  assay.

# **DPPH Radical Scavenging Assay**

DPPH radical scavenging activity the methanol extracts from both parts of the plant was evaluated by the method of Ramasamy *et al.* (2022). A 0.004% DPPH solution was prepared in 95% methanol, and varying volumes (0.1 mL to 0.5 mL) of 10 mg/mL of each extract stock solution were tested. The DPPH solution was introduced, and the absorbance was recorded at 517 nm after 10 minutes. Ascorbic acid was used as the reference standard, and the DPPH free radical scavenging activity was evaluated and quantified using the equation below:

% of DPPH radical scavenging activity  $= \frac{\text{Absorbance of control} - \text{Absorbance of test sample } b}{\text{Absorbance of control}} \times 100$ 

# Molybdenum Assay

The antioxidant activity of the extracts from both the leaves and bark was evaluated using the method described by Wan et al. (2017). 0.5 mL of an aliquot in each test sample was thoroughly mixed with A total of 4.5 mL of reagent (consisting of 28 mM sodium phosphate, 4 mM ammonium molybdate, and 0.6 M sulfuric acid) was combined with 0.5 mL of 45% ethanol, which served as the blank. The resulting mixture was incubated at 95 °C for 90 minutes, after which it was allowed to cool to room temperature. Absorbance was recorded at a wavelength of 695 nm with a UV-2450 spectrophotometer (Shimadzu, Japan). Higher absorbance indicated greater antioxidant activity.

#### **Reducing Power Assay**

The antioxidant enzyme-reducing capability of Fe<sup>3+</sup> in the extracts from bark and leaves was examined as described by Irshad *et al.* (2012). 0.75 mL of the plant extracts at various concentrations was combined with the 0.75 mL phosphate buffer solution (0.2 m and pH 6.6). Then 0.75 mL of a 1% (w/v) potassium hexacyanoferrate (K<sub>3</sub>Fe (CN)<sub>6</sub>) solution. Both solutions were properly combined and incubated for twenty minutes at a temperature of 50 °C. To this mixture,

0.75 mL of a 10 percent TCA solution was added, followed by a combination of 1.5 mL of double distilled water and 0.1 mL of FeCl<sub>3</sub> solution at a concentration of 0.1% (W/V). After ten minutes the absorbance was taken at 700 nm. The highest absorbance indicated a high reducing power activity.

# **Nitric Oxide Scavenging Assay**

The nitric oxide radical scavenging activity of TEMPOfunctionalized dendrimers was assessed utilizing the method described by Ali et al. (2020) involving sodium nitroprusside. For this assay, a mixture was prepared consisting of 2 mL of a ten mM sodium nitroprusside solution and 0.5 mL of phosphate buffer (pH 7.4), combined with 0.5 mL of either TEMPOfunctionalized dendrimers, vitamin C, or a 4-hydroxy TEMPO solution, and incubated at 25 °C for 150 minutes. Following this, 0.5 mL of nitrite was mixed with 1 mL of 0.33% sulfanilic acid (C<sub>6</sub>H<sub>7</sub>NO<sub>3</sub>S) in 2% glacial acetic acid and allowed to react for 5 minutes. Afterward, 1 mL of 1% Naphthyl Ethylenediamine Dihydrochloride (NEDD) was introduced, and the reaction mixture was incubated at 25 °C for 30 minutes. The absorbance results were indicated by a pink solution measured at 540 nm. The percentage of nitric oxide inhibition was calculated using the relevant equation.

Percentage (%) of nitric oxide radical scavenging assay = [(A0-A1)/A0] X 100; where A0 was control, and A1 was absorbance of test sample.

# Hydrogen Peroxide (H,O,) Scavenging Assay

The capability of the extracts to scavenge  $H_2O_2$  was evaluated using a modified approach based on Banothua *et al.* (2017). A 40 mM  $H_2O_2$  solution was created with a 0.05 M phosphate buffer solution (pH7.4). Different concentrations of plant extracts (25, 50, 75, 100 and 125  $\mu$ g/mL) were thoroughly combined with 0.6 mL of the  $H_2O_2$  solution. After a ten-minute interval, the absorbance was measured at 230 nm using a UV-visible spectrophotometer (Shimadzu, Japan), with phosphate buffer without  $H_2O_2$  serving as a blank. The percentage inhibition and  $IC_{50}$  values were determined by using the following equation.

Radical scavenging activity =  $[(OD_{control} - OD_{sample})/OD_{control}]$  X 100, where optical density is the absorbance of samples.

# **Statistical Analysis**

The results were analyzed statistically by using Statistica13.3 software. One-way analysis of variance (ANOVA) with Turkey's post hoc test was used to determine significant differences between the mean values. Values of p<0.05 were statistically significant.

#### **RESULTS AND DISCUSSION**

Recently, people have sought to manage health issues or reduce hazards by improving their diets. Plants and fruits contain a variety of phytochemical constituents with significant antioxidant potential. Natural antioxidants are regarded as effective and promising substitutes for synthetic antioxidants in minimizing oxidation within intricate food systems (Wang et al., 2009). Phytochemical evaluations of plant extracts have identified the existence of phytoconstituents that demonstrate therapeutic and physiological effects (Banothua et al., 2017). In this study, an initial phytochemical analysis was conducted on both the leaf and bark extracts of C. glauca. In leaf extract, methanol is a superior solvent for extracting a diverse range of bioactive compounds compared to chloroform. A higher concentration of flavonoids, phenolics, and proteins was observed in the methanol extract, not only this, alkaloids, glycosides, terpenoids, tannins, coumarins, and amino acids were observed. This finding aligns with several previous studies that have examined the efficacy of methanol solvent in extracting valuable phytochemicals. For instance, Farswan et al. (2009) highlighted the broad spectrum of therapeutic compounds found in C. glauca extracts, emphasizing the importance of selecting appropriate solvents for effective extraction. Similarly Srinivas and Hosamath (2019) also observed the highest phytocompounds from the methanol extract of *C. glauca*.

In the chloroform extract of the leaf, alkaloids, flavonoids, glycosides, and phenolics were detected, but generally at lower concentrations. Flavonoids were present moderately (++), while phenolics were detected in smaller amounts (+). Sugars, tannins, terpenoids, and coumarins were absent, and proteins and amino acids were present at lower levels (Table 1). This represents a lower count of phytocompounds compared to the methanol extract. This finding is consistent with (Tiwari et al., 2023), who also observed a higher count of phytocompounds in methanol extracts compared to chloroform. Additionally, Fraga-Corral et al. (2020) highlighted that methanol's polar nature makes it more effective in extracting a wider range of phytocompounds.

The preliminary phytochemical screening of *C. glauca* bark extracts reveals that methanol is more effective than chloroform for extracting a diverse range of phytochemicals. Methanol consistently yielded higher concentrations of flavonoids, glycosides, phenolics, tannins, and terpenoids compared to chloroform. Alkaloids and proteins were present in both solvents, but flavonoids and glycosides were detected only in the methanol extract. Proteins and amino acids were present in

both extracts, with methanol showing a higher concentration of proteins (++), while both extracts had similar concentrations of amino acids (+). To best of our knowledge there was no previous report on phytochemical composition of *C. glauca* bark extract. The absence of sugars in both extracts suggests either their minimal concentration or limitations in the detection methods used and low-concentration sugars in phytochemical screenings. The commonly utilized term antioxidant capacity refers to a measure used to describe samples obtained from mammals, food, beverages, plant health, and their constituents, which assess their capability to scavenge or neutralize free radicals (Amby *et al.*, 2025).

The preliminary phytochemical screening of C. glauca leaf and bark extracts underscores methanol's superiority over chloroform in extracting a broader range of bioactive compounds. Methanol extracts were rich in flavonoids, phenolics, proteins, alkaloids, glycosides, terpenoids, tannins, and amino acids. They exhibit significant antioxidant and anti-inflammatory characteristics. Potential antimicrobial and anticancer effects, and support cardiovascular health and overall physiological functions (Tungmunnithum et al., 2018; Zandavar & Afshari Babazad, 2023; Bhavikatti et al., 2024). This aligns with the work (Farswan et al., 2009; Srinivas & Hosamath, 2019), who emphasized methanol's effectiveness in extracting diverse phytochemicals. Elevated concentrations of flavonoids, phenolics, and terpenoids indicate notable antioxidant, antiinflammatory, and antimicrobial characteristics. Singh and Tiwari (2022) and Corral et al. (2020) further support these findings, noting methanol's polar nature as advantageous for extracting a wide array of therapeutic compounds. This research examined and compared the antioxidant properties and metabolite profiles of grapes from various regions and types to investigate the possible antioxidant mechanisms of flavonoid metabolites (Fei et al., 2025).

#### Quantitative Analysis of Phytochemicals on Bark Extract

The analysis of the phytochemicals in the bark extract of *C. glauca* shows considerable amounts of phenolics, flavonoids, and tannins. The total phenolic content is measured at 49.259±0.6547 mg of gallic acid equivalents (GAE) per gram of extract, indicating a strong presence of phenolic compounds recognized for their antioxidant properties. These phenolics

Table 1: Preliminary phytochemical analysis on leaf and bark extracts of *Cassine glauca*.

| S. No. | Phytochemical constituents | Leaf             |                    | Bark             |                    |
|--------|----------------------------|------------------|--------------------|------------------|--------------------|
|        |                            | Methanol Extract | Chloroform Extract | Methanol Extract | Chloroform Extract |
| 1      | Flavonoids                 | +++              | ++                 | +                | -                  |
| 2      | Alkaloids                  | +                | +                  | ++               | ++                 |
| 3      | Phenolics                  | ++               | +                  | ++               | +                  |
| 4      | Glycosides                 | +                | +                  | +                | -                  |
| 6      | Tannins                    | ++               | -                  | +++              | +                  |
| 7      | Coumarins                  | +                | -                  | -                | -                  |
| 8      | Sugars                     | -                | -                  | -                | -                  |
| 10     | Proteins                   | +++              | -                  | ++               | +                  |
| 11     | Terpenoids                 | +                | -                  | +++              | ++                 |
| 12     | Amino acids                | ++               | +                  | +                | +                  |

(+) low; (++) average; (+++) high; (-) absent

Table 2: Quantitative analysis of phenolics, flavonoids and tannins of bark and leaf extracts of C. glauca.

| S. No. | Analytical parameters | Phytochemicals quantity in bark extract (mg) | Phytochemicals quantity in leaf extract (mg)   |
|--------|-----------------------|--|--|
| 1      | Total phenolics       | 49.259±0.6547 GAE <sup>a</sup> /g extract    | 47.458±0.5748 GAE <sup>a</sup> /g extract      |
| 2      | Total flavonoids      | 54.24±0.5478 TAEb/g extract                  | $160.106\pm0.3014$ TAE <sup>b</sup> /g extract |
| 3      | Total tannins         | 140.307±0.6547 RE <sup>c</sup> /g extract    | 45.298±0.9874 REc/g extract                    |

GAE<sup>a</sup> - gallic acid equivalents, TAE<sup>b</sup> - Tannic acid equivalents, RE<sup>c</sup> - rutin equivalents

enhance the antioxidant capacity of the extract, playing a vital role in neutralizing free radicals and possibly reducing damage associated with oxidative stress.

The total flavonoid content is measured at 54.24±0.5478 mg of tannic acid equivalents (TAE) for each gram of extract, as shown in Table 2. This indicates a significant concentration of flavonoids in the extract. Flavonoids are known for their various biological activities, including anti-inflammatory and antioxidant effects, which further emphasize the potential therapeutic advantages of the extract. Additionally, the total tannin content stands at 140.307±0.6547 mg of rutin equivalents (RE) per gram of extract. According to estimates, approximately 50,000 to 70,000 plant species possess medicinal properties, emphasizing their vital importance in both traditional and contemporary healthcare systems (Ali et al., 2025).

# Quantitative Analysis of Phytochemicals on Leaf Extract

The analysis of the leaf extract from *C. glauca* shows considerable amounts of phenolics, flavonoids, and tannins. The overall phenolic content stands at 47.458±0.5748 mg equivalent to gallic acid (GAE) per gram of extract, highlighting a significant presence of phenolic compounds renowned for their antioxidant effects. Phenolics boost the antioxidant capacity of the extract, aiding in the neutralization of free radicals and potentially mitigating damage associated with oxidative stress. The total flavonoid content measures 160.106±0.3014 mg equivalent to tannic acid (TAE) per gram of extract and highlighting a rich concentration of flavonoids. These compounds are renowned for their antioxidants, biological activities, and anti-inflammatory effects, supporting the extract's potential therapeutic benefits. The total tannin content is  $45.298 \pm 0.9874$  mg rutin equivalents (RE) per gram of extract Table 2. While lower compared to phenolics and flavonoids, this substantial concentration of tannins is associated with various health benefits, including antimicrobial and antioxidant activities (Muniyandi et al., 2019; Sundari et al., 2024). Veerapur et al. (2017) performed a quantitative analysis of flavonoids in C. glauca. In contrast, the earlier study identified various phytocompounds, including flavonoids and phenolics, in C. glauca. According to the literature review, there has been no quantitative analysis of phytochemicals in the bark and leaf extracts of C. glauca (Faruq et al., 2024; Adugna et al., 2024). Key phytochemicals that demonstrate significant efficacy against these mites include phenol, flavonoids, terpenoids, tannins, and phenylpropanoids. They exert their effects by inducing neurotoxicity, disrupting digestion and metabolism, damaging the cuticle, causing oxidative stress, harming cell membranes, inhibiting respiration, and disrupting hormonal balance in these mites (Mustafa & Alsayegh, 2025).

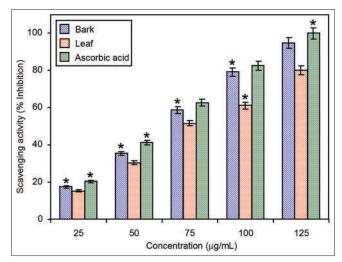


Figure 1: The DPPH assay of Cassine glauca bark and leaf extract at different concentrations

In the present study, the subsequent section of the analysis was the antioxidant activity of C. glauca leaf and bark extracts. Based on the phytochemical study, the highest count of phytochemicals containing methanol extract was selected for antioxidant activity with 5 different methods. The antioxidant properties of extracts from C. glauca were assessed and compared with ascorbic acid, which is a standard antioxidant, across various concentrations. The results revealed significant differences in antioxidant activity among the tested materials. At the lowest concentration of 25  $\mu$ g/mL, ascorbic acid demonstrated the highest antioxidant activity with 20.4% inhibition, compared to 17.4% for the bark extract and 15.3% for the leaf extract. Finally, at 125  $\mu$ g/mL, the bark extract reached 94.7% inhibition, surpassing the leaf extract's 80.3%, but ascorbic acid remained the most effective with 100% inhibition (Figure 1).

These findings suggest that while both *C. glauca* extracts exhibit strong antioxidant activity, the bark extract demonstrates particularly high activity at higher concentrations. Ascorbic acid, however, remains the most effective antioxidant across all concentrations. This is consistent with previous studies that have shown ascorbic acid's potent antioxidant properties due to its capacity to counteract free radicals and diminish oxidative stress (Bhattarcharya *et al.*, 2018). Additionally, research by El-Hashash *et al.* (2010) highlighted the effectiveness of various species of *Cassine* extracts. The increased activity of both extracts at elevated concentrations suggests that they could serve as an important source of antioxidants, aligning with findings from another study that emphasize the therapeutic potential of *C. glauca* (Kumar *et al.*, 2013; El-Hashash *et al.*, 2010).

The molybdenum assay measures antioxidant capacity by mixing a sample with a reagent containing molybdenum (VI) ions. Antioxidants in the sample reduce molybdenum (VI) to molybdenum (V), forming a green complex with phosphoric acid. This green color, quantified via spectrophotometry, reflects the sample's antioxidant activity. The assay thus gauges the sample's ability to neutralize oxidative stress. In the present study, using at 25 µg/mL of the bark extract exhibited the lowest activity at 3.3%, while the leaf extract showed 11.7%, and ascorbic acid demonstrated the highest activity at 15.6%. At 125 µg/mL, the bark extract reached its highest activity at 28.7%, the leaf extract showed 51.8%, and ascorbic acid remained the most effective with 71.4% (Figure 2). These results highlight that while both C. glauca extracts display increasing antioxidant activity with concentration, the leaf extract consistently outperforms the bark extract, and ascorbic acid maintains superior antioxidant activity across all tested concentrations. The findings are consistent with previous research that underscores the antioxidant potential of C. glauca. Based on the literature review, there is limited research on antioxidant determination using the molybdenum assay. El-Hashash et al. (2010) examined the antioxidant properties of different Cassine species utilizing this approach.

The reducing power is frequently utilized as a measure of electron-donating capability, a crucial method for assessing the antioxidative effects, antioxidant properties, or radical scavenging potential of the extracts. The reducing power of *C. glauca* extracts was evaluated and compared to ascorbic acid. At 25 µg/mL, the bark extract had a reducing power of 7.4%, the leaf extract 11.2%, and ascorbic acid 12.4%. At 50 µg/mL, the values increased to 12.3% for the bark extract, 19.4% for the leaf extract, and 25.4% for ascorbic acid. Finally, at 125 µg/mL, the bark extract showed 52.7%, the leaf extracts 58.2%, and the ascorbic acid 68.7% (Figure 3). These results reveal that while both *C. glauca* extracts showed that increasing reducing power with concentration, ascorbic acid consistently remains the most effective. To date, no studies have investigated the antioxidant activity of *C. glauca* using the reducing power method. The

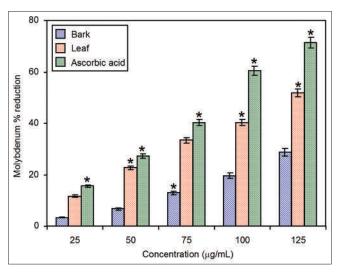


Figure 2: The percentage of molybdenum scavenging activity of Cassine glauca bark and leaf extract

presented study correlated with estimation of antioxidants in *Benkara malabarica* (Kalusalingam & Balakrishnan, 2022b).

The nitric oxide (NO) assay evaluates antioxidant activity by measuring the ability to scavenge nitric oxide radicals, which are involved in various processes. In this assay, antioxidants inhibit the formation of nitrite from nitric oxide, reflecting their potential. In the present study, the next antioxidant method was the Nitric oxide method, at 25 µg/mL, the bark extract showed 4.7% inhibition of nitric oxide, while the leaf extract exhibited 6.9%, and ascorbic acid demonstrated a higher inhibition of 8.3%. At 100 μg/mL, the bark extract reached 27.8%, the leaf extract 35.7%, and ascorbic acid 36.7%. Finally, at 125 μg/mL, the bark extract showed 45.3% inhibition, the leaf extracts 49.8%, and the ascorbic acid 52.5% (Figure 4). These results indicate that both C. glauca extracts demonstrate increasing antioxidant activity with concentration, with the leaf extract consistently showing higher activity compared to the bark extract. Ascorbic acid remains the most effective inhibitor of

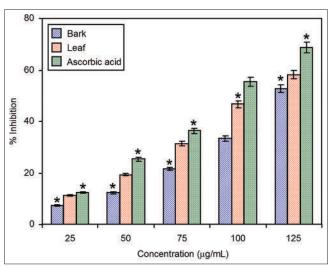


Figure 3: The reducing power assay of *Cassine glauca* bark and leaf extract at different concentrations

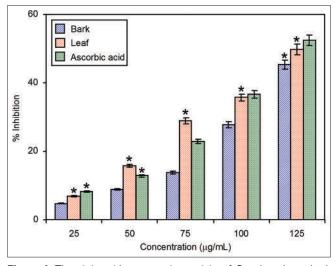


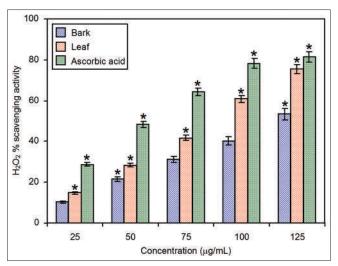
Figure 4: The nitric oxide scavenging activity of Cassine glauca bark and leaf extract at different concentrations

nitric oxide across all concentrations. Both *C. glauca* extracts demonstrated increased inhibition with higher concentrations, with the leaf extract showing superior activity compared to the bark extract.

The antioxidant properties of C. glauca extracts were evaluated using the hydrogen peroxide  $(H_2O_2)$  assay and compared to ascorbic acid. At 25 µg/mL, the bark extract exhibited 10.3% inhibition of  $H_2O_2$ , while the leaf extract showed 14.7%, and ascorbic acid demonstrated a higher inhibition of 28.7%. Finally, at 125 µg/mL, the bark extract demonstrated 53.4% inhibition, the leaf extracts 75.4%, and ascorbic acid 81.4% (Figure 5). These results indicate that both C. glauca extracts showed increasing antioxidant activity with concentration, with the leaf extract consistently exhibiting higher activity compared to the bark extract. Ascorbic acid remained the most effective inhibitor of  $H_2O_3$  across all concentrations.

These findings validate the  $H_2O_2$  assay as a reliable technique for evaluating the antioxidant potential of natural extracts, as it effectively measures the ability of antioxidants to mitigate oxidative stress by neutralizing hydrogen peroxide. The observed trends are consistent with previous research that highlights the antioxidant properties of C. glauca and confirms the effectiveness of the  $H_2O_2$  assay in assessing the antioxidant activity of various substances. This technique is both efficient and highly accurate, making it ideal for the quick measurement of  $H_2O_2$  scavenging capacity in standard antioxidants as well as in natural antioxidants found in plant extracts (Fernando & Soysa, 2015).

Overall, the study confirms that both *C. glauca* extracts exhibit significant antioxidant activity, with methanol extracts demonstrating superior efficacy, because of their greater concentration of bioactive substances. The leaf extract generally shows better performance than the bark extract in antioxidant assays, while ascorbic acid remains the most potent antioxidant across all tested methods. Among the five methods used, the DPPH assay consistently provided



**Figure 5:** The Hydrogen Peroxide scavenging assay of *Cassine glauca* bark and leaf extract at different concentrations.

the highest antioxidant activity, making it the most effective for evaluating antioxidant potential. Its sensitivity to radical scavenging activity resulted in the highest inhibition levels, underscoring its effectiveness compared to the other methods. A variety of new compounds including oxylipins, lignans, saponins, flavonoids, phenolics, and their derivatives have been identified. This highlights the plant's potential as a medicinal resource within alternative medicine systems such as Ayurveda, Siddha, and Unani for addressing various metabolic disorders, enabling the plant's roots to demonstrate a range of pharmacological effects like regulating glucose and fatty acid cycles, along with antioxidant and anti-inflammatory properties (Devadasu & Martin, 2025).

#### **CONCLUSION**

The phytochemical analysis of C. glauca extracts from both leaf and bark reveals considerable quantities of bioactive ingredients, such as phenolics, flavonoids and tannins, along with the leaf extract showing higher concentrations than the bark extract. Methanol extraction proves to be more effective than chloroform, highlighting its superiority in obtaining these valuable compounds. The high levels of flavonoids, phenolics and tannins underscore the extracts' potential as natural antioxidants and anti-inflammatory substances, which are essential for reducing oxidative stress and inflammation linked to chronic conditions like cardiovascular diseases, cancer, and neurodegenerative conditions. The remarkable antioxidant properties of the leaf extract bolster its promise as a treatment for disorders associated with oxidative stress. The research suggests that both leaf and bark extracts possess significant pharmaceutical potential, necessitating additional studies to isolate and identify the specific bioactive compounds and investigate their mechanisms of action and clinical uses.

## **ACKNOWLEDGMENT**

The authors are thankful to the Department of Botany, Arignar Anna Government Arts College, Namakkal for providing support for experimenting. The authors are also thankful to Dr.M.Paulpandi, Bharthiar University, Coimbatore for interpretation of antioxidant analysis of the samples.

## **REFERENCES**

Adugna, H., Ezez, D., Guadie, A., & Tefera, M. (2024). Phytochemical profiling, evaluation of heavy metal load, antioxidant and antibacterial activity of various medicinal plants. *Journal of Agriculture and Food Research, 16*, 101190. https://doi.org/10.1016/j.jafr.2024.101190

Albrahim, J. S., Alosaimi, J. S., Altaher, A. M., Almulayfi, R. N., & Alharbi N. F. (2021). Employment of *Cassia angustifolia* leaf extract for Zinc Nanoparticles fabrication and their antibacterial and cytotoxicity. *Saudi Journal of Biological Sciences, 28*(6), 3303-3308. https://doi.org/10.1016/j.sjbs.2021.02.075

Ali, B. M., Boothapandi, M., & Nasar, A. S. (2020). Nitric oxide, DPPH and hydrogen peroxide radical scavenging activity of TEMPO terminated polyurethane dendrimers: Data supporting the antioxidant activity of radical dendrimers. Data in Brief, 28, 104972. https://doi. org/10.1016/j.dib.2019.104972

- Ali, S., Ali, H., Hussain, A., Dey, G., Wang, C.-W., Taharia, M., Lin, P.-Y., Sharma, R. K., Hsiao, P.-G., & Chen, C. Y. (2025). Comparative analysis of phytochemical and antioxidant potential in traditional medicinal plants: Insights from solvent-based extractions. *Food Bioscience*, 68, 106603. https://doi.org/10.1016/j.fbio.2025.106603
- Amby, D. B., Mekureyaw, M. F., Akhtar, S. S., Pandey, C., & Roitsch, T. (2025). Simple and semi-high throughput determination of total phenolic, anthocyanin, flavonoid content, and total antioxidant capacity of model and crop plants for cell physiological phenotyping. *Plant Science*, 357, 112524. https://doi.org/10.1016/j. plantsci.2025.112524
- Banothua, V., Neelagiria, C., Adepallya, U., Lingamb, J., & Bommareddy, K. (2017). Phytochemical screening and evaluation of *in vitro* antioxidant and antimicrobial activities of the indigenous medicinal plant *Albizia odoratissima*. *Pharmaceutical Biology, 55*(1), 1155. https://doi.org/10.1080/13880209.2017.1291694
- Bhattarcharya, A., Tiwari, P., Sahu, P. K., & Kumar, S. (2018). A Review of the phytochemical and pharmacological characteristics of *Moringa oleifera*. *Journal of Pharmacy and Bioallied Sciences*, *10*(4), 181-191. https://doi.org/10.4103/JPBS.JPBS\_126\_18
- Bhavikatti, S. K., Zainuddin, S. L. A., Ramli, R. B., Nadaf, S. J., Dandge, P. B., Khalate, M., & Karobari, M. I. (2024). Insights into the antioxidant, anti-inflammatory and anti-microbial potential of *Nigella sativa* essential oil against oral pathogens. *Scientific Reports*, 14, 11878. https://doi.org/10.1038/s41598-024-62915-1
- Calixto, J. B. (2000). Efficacy, safety, quality control, marketing, and regulatory guidelines for herbal medicines (phytotherapeutic agents). Brazilian Journal of Medical and Biological Research, 33(2), 179. https://doi.org/10.1590/S0100-879X2000000200004
- Chintamunnee, V., & Mahomoodally, M. F. (2012). Herbal medicine is commonly used against non-communicable diseases in the tropical island of Mauritius. *Journal of Herbal Medicine*, *2*(4), 113-125. https://doi.org/10.1016/j.hermed.2012.06.001
- Devadasu, V., & Martin, A. (2025). Comprehensive assessment of the nutritional, phytochemical, and volatile components present in the roots of *Asparagus racemosus*, an underutilized plant for food applications. *Food Bioscience*, 68, 106677. https://doi.org/10.1016/j. fbio.2025.106677
- Dhanasekaran, K., Kishore kumar, A., & Balakrishnan, V. (2025). Antibacterial and antifungal activities from leaf and bark extract of *Cassine glauca* (Rottb.) Kuntze. *Current Botany, 16*, 53-58. https://doi.org/10.25081/cb.2025.v16.9510
- El-Hashash, M. M., Abdel-Gawad, M. M., El-Sayed, M. M., Sabry, W. A., Abdel-Hameed, E. S., & Abdel-Lateef, E. S. (2010). Antioxidant properties of methanolic extracts of the leaves of seven Egyptian Cassia species. Acta Pharmaceutica, 60, 361-367. https://doi. org/10.2478/v10007-010-0030-y
- Farswan, M., Mazumder, P. M., & Percha, V. (2009). Protective effect of Cassia glauca Linn. on the serum glucose and hepatic enzymes level in streptozotocin induced NIDDM in rats. *Indian Journal of Pharmacology*, 41(1), 19-22. https://doi.org/10.4103/0253-7613.48887
- Faruq, M. O., Rahim, A., Arifuzzaman, M., & Ghosh, G. P. (2024). Phytochemicals screening, nutritional assessment and antioxidant activities of A. viridis L. and A. spinosus L. leaves A comparative study. *Journal of Agriculture and Food Research*, 18, 101341. https://doi.org/10.1016/j.jafr.2024.101341
- Fei, L., Zhang, D., Mao, Y., Mkunga, J. J., Chen, P., He, C., Shan, C., Yang, X., & Cai, W. (2025). Metabolomics combined with network pharmacology reveals the regional and variety heterogeneity of grape metabolites and their potential antioxidant mechanisms. Food Research International, 211, 116443. https://doi.org/10.1016/j. foodres.2025.116443
- Fernando, C. D., & Soysa, P. (2015). Optimized enzymatic colorimetric assay for determination of hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) scavenging activity of plant extracts. *MethodsX*, 2, 283-291. https://doi.org/10.1016/j.mex.2015.05.001
- Fraga-Corral, M., García-Oliveira, P., Pereira, A. G., Lourenço-Lopes, C., Jimenez-Lopez, C., Prieto, M. A., & Simal-Gandara, J. (2020). Technological Application of Tannin-Based Extracts. *Molecules*, 25(3), 614. https://doi.org/10.3390/molecules25030614
- Harborne, J. B. (1973). *Phytochemical Methods: A Guide to Modern Techniques of Plant Analysis*. London, UK: Chapman and Hall.
- Hassan, B. A. R. (2012). GC-MS analysis, *in-vitro* anti-diabetic and anti-oxidant activities of *Rhynchosia nummularia* (L.) DC. *Pharmaceitica Analytica*

- Acta, 3(10), 139. https://doi.org/10.4172/2153-2435.1000e139
- Irshad, M., Zafaryab, M., Singh, M., & Rizvi, M. M. A. (2012). Comparative analysis of the antioxidant activity of *Cassia fistula* extracts. *International Journal of Medicinal Chemistry, 1*, 1-6. https://doi.org/10.1155/2012/157125
- Iyiola, A. O., & Wahab, M. K. A. (2023). Herbal Medicine Methods and Practices in Nigeria. In S. C. Izah, M. C. Ogwu, & M. Akram (Eds.), Herbal Medicine Phytochemistry (pp. 1395-1428) Cham, Switzerland: Springer. https://doi.org/10.1007/978-3-031-43199-9\_47
- Kalusalingam, M., & Balakrishnan, V. (2022a). Phytochemical, antimicrobial and antioxidant analysis of indigenously used folk medicinal plant *Ixora notoniana* Wall. *Current Trends in Biotechnology and Pharmacy,* 16(1), 64-76. https://doi.org/10.5530/ctbp.2022.1.7
- Kalusalingam, M., & Balakrishnan, V. (2022b). Estimation of antioxidant and cytotoxicity activities of extracts obtained from the leaves of Folk medicinal plant Benkara malabarica (Lam). Indian Journal of Pharmaceutical Education and Research, 56(3), 838-845.
- Kavitha, R., Sundari, T., & Srinivasan P. (2024). Chemoprofiling of *Cucumis pubescens* Willd. fruits. *Journal of Phytology*, 16, 127-132. https://doi.org/10.25081/jp.2024.v16.8938
- Kokate, C. K. (2005). A Textbook for Practical Pharmacognosy. (5th ed). New Delhi, India: Vallabh Prakashan.
- Mosihuzzaman, M. (2012). Herbal Medicine in Healthcare-An Overview. *Natural Product Communicatins*, 7, 807. https://doi.org/10.1177/1934578X1200700628
- Muniyandi, K., George, E., Sathyanarayanan, S., George, B. P., Abrahamse, H., Thamburaj, S., & Thangaraj, P. (2019). Phenolics, tannins, flavonoids and anthocyanins contents influenced antioxidant and anticancer activities of Rubus fruits from Western Ghats, India. Food Science and Human Wellness, 8(1), 73-81. https://doi. org/10.1016/j.fshw.2019.03.005
- Muscolo, A., Mariateresa, O., Giulio, T., & Russo, M. (2024). Oxidative Stress: The Role of Antioxidant Phytochemicals in the Prevention and Treatment of Diseases. *International Journal of Molecular Sciences*, 25(6), 3264. https://doi.org/10.3390/ijms25063264
- Mustafa, S., & Alsayeqh, A. F. (2025). Role of plant phytochemicals/ extracts for the control of *Dermanyssus gallinae* in poultry and its zoonotic importance. *Poultry Science*, *104*(4), 104899. https://doi. org/10.1016/j.psj.2025.104899
- Raina, H., Soni, G. Jauhari, N., Sharma, N., & Bharadvaja, N. (2014). Electron micrographic representations of mechanisms of action of murine Norovirus on ATCC TIB-71 Cells and level of gene expression. *Turkish Journal of Botany*, 38(6), 1027. https://doi.org/10.3906/bot-1405-93
- Ramasamy, R., Murugesh, S., Jegadeesh Kumar, D., Prakash, B., Kasirajan, G., & Gayathri, G. (2022). Characterization and evaluation of antimicrobial, antioxidant and antibiofilm activities of silver nanoparticles biosynthesised from *Harpullia arborea* bark extract. *Journal of Clinical and Diagnostic Research*, 16(9), FC07. https://doi.org/10.7860/JCDR/2022/56639.16859
- Sreelatha, S., & Padma, P. R. (2009). Antioxidant activity and total phenolic content of *Moringa oleifera* leaves in two stages of maturity. *Plant Foods for Human Nutrition, 64*, 303-311. https://doi.org/10.1007/s11130-009-0141-0
- Srinivas, Y., & Hosamath, P. (2019). Phytochemical and *in vitro* anticancer activity of *Cassia glauca* leaves extract. *International Journal of Green Pharmacy*, 13(4), 354. https://doi.org/10.22377/ijgp.v13i04.2707
- Sundari, T., & Kavitha, R. (2024). *In vitro* assessing of *Cucumis pubescens* Willd. fruit extract for phytochemical, antibacterial, antioxidants and toxicity assays. *Journal of the Indian Chemical Society, 101*(7), 101176. https://doi.org/10.1016/j.jics.2024.101176
- Sundari, T., Kavitha, R., & Aarthi, M. (2024). *In silico* analysis of *Cucumis* pubescens Willd. Fruit extract phytocompounds and its activity against anti-diabetic targets. *Notulae Scientia Biologicae*, 16(3), 12049.
- Tiwari, H., Singh, P. K., Naresh, R. K., Ismayil, M. M. S., Monika, S., Islam, A., Kumar, S., Singh, K. V., Pandey, A. K., & Shukla, A. (2023). Millets based integrated farming system for food and nutritional security, constraints and agro-diversification strategies to fight global hidden hunger: A Review. *International Journal of Plant & Soil Science*, 35(19), 630-643. https://doi.org/10.9734/ijpss/2023/v35i193593
- Tungmunnithum, D., Thongboonyou, A., Pholboon, A., & Yangsabai, A. (2018). Flavonoids and Other Phenolic Compounds from Medicinal Plants for Pharmaceutical and Medical Aspects: An Overview.

- Medicines, 5(3), 93. https://doi.org/10.3390/medicines5030093
- Veerapur, V. P., Pratap, V., Thippeswamy, B. S., Marietta, P., Bansal, P., Kulkarni, P. V., & Kulkarni, V. H. (2017). Aldose reductase inhibitor form Cassia glauca: A comparative study of cytotoxic activity with Ag nanoparticles (NPs) and molecular docking evaluation. Journal of Ethnopharmacology, 198, 489-498. https://doi.org/10.1016/j. jep.2017.01.025
- Wan, C., Yu, Y., Zhou, S., Liu, W., Tian, S., & Cao, S. (2017). Antioxidant activity and free radical-scavenging capacity of Gynuradi varicata leaf extracts at different temperatures. *Pharmacognosy Magazine*, 7(25), 4-45. https://doi.org/10.4103/0973-1296.75900
- Wang, L., Zhang, X.-Q., Yin, Z.-Q., Wang, Y., & Ye, W.-C. (2009). Two new Amaryllidaceae alkaloids from the bulbs of *Lycoris radiata*. *Chemical and Pharmaceutical Bulletin, 57*(6), 610. https://doi.org/10.1248/cpb.57.610
- Xu, D., Hu, M.-J., Wang, Y.-Q., & Cui, Y.-L. (2019). Antioxidant activities of Quercetin and its complexes for medicinal application. *Molecules*, 24(6), 1123. https://doi.org/10.3390/molecules24061123
- Zandavar, H., & Afshari Babazad, M. (2023). Secondary Metabolites: Alkaloids and Flavonoids in Medicinal Plants. In E. Ivanišová (Eds.), Herbs and Spices-New Advances (pp. 1-10) London, UK: IntechOpen Limited. https://doi.org/10.5772/intechopen.108030